Nanomaterial Sorbents for the Preparation of ¹⁸⁸W/¹⁸⁸Re Generator and ¹⁸⁸Re Radiopharmaceutical Development

Madhava B. Mallia¹, Rubel Chakravarty¹, Ashutosh Dash^{1,*} and Maroor Raghavan Ambikalmajan Pillai²

¹Radiopharmaceuticals Division, Bhabha Atomic Research Centre, Mumbai-400085, India

²Molecular Group of Companies, Puthuvype, Ernakulam-682508, India

Abstract: Rhenium-188 is a useful radionuclide for targeted therapy thanks to the high energy (2.12 MeV) β^{-} emission, low abundance gamma rays (155 keV and 15%) suitable for imaging and availability from the ¹⁸⁸W/¹⁸⁹Re generator. Though considerable amount of work on the development of ¹⁸⁸Re-radiopharmaceuticals have been reported in the literature, the clinical uses of them are still low mainly due to the limited availability and high cost of the generators. However, a ¹⁸⁸Re radiopharmaccy can be successfully run provided the generator is used for the preparation of several ¹⁸⁸Re-radiopharmaceuticals. The relatively low radioactive concentration of ¹⁸⁹Re from conventional alumina based generators, thereby needing post elution concentration, is one of the limitations for the extended use of the generator. The development of nanomaterials having higher capacity for adsorption of ¹⁸⁸W allows it to be used for the preparation of course of column generators capable of giving high radioactive concentration for the eluted perrhenate. This review provides an overview of the work done at the Bhabha Atomic Research Centre, Mumbai, India, for the development of ¹⁸⁸Re/service.

Keywords: Nanomaterials, Radionuclide generator, Radiopharmaceuticals, Rhenium-188, Targeted therapy.

1. INTRODUCTION

Rhenium belongs to the group 7 of the periodic table, same as that of 99mTc, the workhorse of nuclear medicine. Rhenium has two important radioisotopes, ^{186}Re [half-life – 3.71 days, E_{Bmax} – 1.09 MeV, E_{y} - 136 keV (9%)] and ^{188}Re [half-life – 16.9 h, $E_{\beta max}$ – 2.12 MeV, E_v - 155 keV (15%)][1], both of which have beta energies suitable for therapy and gamma emissions, though of low abundance, which permits monitoring the distribution of the radiotracer in vivo, and potentially, perform image-based dosimetry calculations. While relatively low-energy beta particles of ¹⁸⁶Re are useful for therapeutic applications requiring low tissue penetration, high-energy beta from ¹⁸⁸Re are particularly useful for therapy of diseases in large organs such as liver. Being in the same group of the periodic table, technetium and rhenium share similar chemistry, which implies, wherever applicable, it is possible to have theranostic pair with ^{99m}Tc and ^{186/188}Re radionuclides. However, rhenium being more difficult to reduce than technetium, guite often, technetium chemistry cannot be directly extrapolated for preparing rhenium radiopharmaceuticals. Rhenium generally requires harsher radiolabeling conditions compared to technetium, and therefore, ligands that

Address correspondence to this author at the Radiopharmaceuticals Division, Bhabha Atomic Research Centre, Mumbai 400 085, and India; Tel: 91-22-2559 5372; Fax: 91-22-2550 5345; cannot survive these conditions cannot be used for preparing rhenium radiopharmaceuticals.

Rhenium-186 is a reactor-produced radionuclide from neutron activation of enriched ¹⁸⁵Re or natural rhenium. A difficulty associated with the production of ¹⁸⁶Re from natural rhenium is the co-production of ¹⁸⁸Re. Though mixed radiation therapy with ^{186/188}Re is feasible, the approach has not been further pursued. On the other hand, ¹⁸⁸Re is a decay product of long lived ¹⁸⁸W and hence available from a ¹⁸⁸W/¹⁸⁸Re generator. Like the ⁹⁹Mo/^{99m}Tc generator, which has a major role in making ^{99m}Tc the workhorse of nuclear medicine, ¹⁸⁸W/¹⁸⁸Re generator has the potential to popularize the use of ¹⁸⁸Re-radiopharmaceuticals for therapy.

Commercial availability of ¹⁸⁸W/¹⁸⁸Re generator has a significant role to play in popularizing the use of ¹⁸⁸Re-radiopharmaceuticals in nuclear medicine. An equally important aspect is to make available new ¹⁸⁸Re-radiopharmaceuticals and freeze dried kits for their preparation in hospital radiopharmacy housing the ¹⁸⁸W/¹⁸⁸Re generator. In this review, authors have attempted to cover the recent developments in ¹⁸⁸W/¹⁸⁸Re generator technology and discuss some of the important ¹⁸⁸Re-radiopharmaceuticals, which have been used in the clinic or which have clinical potential. These discussions are mainly focused on the developmental activities done at the Bhabha Atomic Research Centre, India.

E-mail: adash@barc.gov.in

2. DISCUSSION

2.1. Nanosorbents for ¹⁸⁸W/¹⁸⁸Re Generator Preparation

¹⁸⁸W/¹⁸⁸Re generator is an excellent source for availing no-carrier-added (NCA) grade ¹⁸⁸Re. Most of the separation methodologies which have been reported for ⁹⁹Mo/^{99m}Tc generators have also been exploited for preparation of ¹⁸⁸W/¹⁸⁸Re generators [2]. Out of these procedures, the alumina based column chromatographic approach wherein ¹⁸⁸W is absorbed on bulk alumina matrix and ¹⁸⁸Re is selectively eluted using saline solution at regular intervals, has been identified as the most reliable method for the preparation of ¹⁸⁸W/¹⁸⁸Re generator. Owing to the limited sorption capacity of bulk alumina (~50 mg W/g), clinical-scale ¹⁸⁸W/¹⁸⁸Re generator can only be prepared using high specific activity (150-190 GBg/g) ¹⁸⁸W that can be produced in only very few high flux (~10¹⁵ n.cm⁻².s⁻¹) reactors available (High Flux Isotope Reactor at Oak Ridge National Laboratory in United States and SM Reactor at Dimitrograd in Russia) in the world. Even while using high specific activity ¹⁸⁸W produced in these reactors, the ¹⁸⁸W/¹⁸⁸Re generators currently available yield low specific volume (activity/ mL) of ¹⁸⁸Re and require post-elution concentration procedures prior to radiopharmaceutical preparation, which is not always very convenient to perform in hospital radiopharmacies [3]. From this perspective, it is desirable to develop a $^{188}W/^{188}Re$ generator where the concentration step can be avoided to simplify the operational procedure for widespread clinical utility.

Use of nanomaterials as adsorbents for ¹⁸⁸W/¹⁸⁸Re generator holds promise as they take advantage of the unique physiochemical properties realized at nanoscale that cannot be anticipated from bulk counterparts of the same chemical composition. The defining feature of a nanomaterial is that its properties depend not only on composition but also on its size and shape. One of the specific properties of this class of material is that a high percent of the atoms lies on the surface and the material possesses high surface area. These surface atoms are unsaturated and possess high chemical activity. Therefore, such materials demonstrate high sorption capacity and enhanced selectivity for ¹⁸⁸W resulting in elution of ¹⁸⁸Re with adequate radioactive concentration and high radionuclidic purity. The above properties make nanomaterials an exciting platform to develop into the realm of ¹⁸⁸W/¹⁸⁸Re generator technology with properties that are difficult to achieve using bulk materials.

The ability of three nanomaterial based sorbents (nanocrystalline titania (TiP), zirconia (ZrO₂) and alumina (y-Al₂O₃)) have been comprehensively studied and profusely explored for the development of ¹⁸⁸W/¹⁸⁸Re generators by our group [4-7]. Taking advantage of the new physical properties uniquely associated with nanomaterials, ¹⁸⁸W/¹⁸⁸Re generators were prepared using ¹⁸⁸W having specific activity of 3-5 Ci/g. A scrutiny of the results depicted in Table 1 reveals that all these sorbents are suitable for preparation of ¹⁸⁸W/¹⁸⁸Re generators. Though testing of these nanomaterials was done by using lower load of ¹⁸⁸W, the capacity of the generator can easily be scaled up to >37 GBq (1 Ci) activity levels as the capacity of the nanomatarials are very high. Among all the sorbents used, y-Al₂O₃ was found to exhibit the highest sorption capacity under dynamic conditions as compared to TiP and ZrO₂ and is therefore the most appropriate sorbent for the preparation of clinical-scale ¹⁸⁸W/¹⁸⁸Re generator. The performance of these generators was studied for more than 6 months and reported to be satisfactory. Not only were the ¹⁸⁸Re elution yields appreciably high but also the level of radionuclidic, radiochemical and chemical impurities in the ¹⁸⁸Re obtained from all these generators were well within the acceptable limits prescribed in the pharmacopoeias [8].

The mechanism of uptake of ¹⁸⁸W and elution of ¹⁸⁸Re from the nanosorbent based chromatographic columns could be explained on the basis of pH dependent surface charge on nanosorbents. There are various anionic species of tungsten depending on the pH of the medium. At pH 6, the predominant species is $W_{12}O_{41}^{10-}$ and between pH 2 and 4 the species W₁₂O₃₉⁶⁻ predominates. Owing to electrostatic attraction, highly negatively charged tungsten anion is taken up on the positively charged surface of the nanosorbent under mildly acidic conditions. Subsequently, it may form a stable complex of the type $[AIW_6O_{24}]^{8-}$, similar to that reported for bulk alumina. As these tungstate ions start transforming into perrhenate ion $(^{188}\text{ReO}_4)$, which has only the mononegative charge, the binding gets weaker and an easy displacement of ¹⁸⁸ReO₄⁻ is expected from the nanosorbent.

The performance of the ¹⁸⁸W/¹⁸⁸Re generators prepared using nanomaterial based sorbents is comparable with conventional alumina based generators in terms of ¹⁸⁸Re elution yield and the purity of the eluate. However, due to higher sorption capacity of nanomaterials, relatively lower specific activity ¹⁸⁸W produced in medium flux research reactors can also be used for preparation of clinical-scale ¹⁸⁸W/¹⁸⁸Re generators while using such materials as the sorbent matrix. It is also pertinent to point out that agglomerated nanomaterials (comprising of nanosized crystallites) are used as sorbents for preparation of radionuclide generators. Typically, the size of the agglomerates is in the range of 50-100 mesh size (149-297 µm), which is comparable to the size of bulk alumina particles used in preparation of conventional generators. Controlled agglomeration of nanoparticles is essential in order to use them as sorbents for radionuclide generators, as fully dispersed nanoparticles would exhibit issues such as increased back pressure during the elution process and might also result in breakthrough of nanoparticles in the ¹⁸⁸Re eluate. While using agglomerated nanomaterial based sorbents, no change in the generator design and the elution procedure is required.

Adoption of nanosorbents (especially γ -Al₂O₃) for preparation of ¹⁸⁸W/¹⁸⁸Re generator would reduce reliance on the two high flux reactors for ¹⁸⁸Wproduction. There are a number of research reactors in the world having thermal neutron flux >5 × 10¹⁴ n cm⁻² s⁻¹ which could be used for the production of ¹⁸⁸W for making ¹⁸⁸W/¹⁸⁸Re generators. Also if the existing generator manufacturers can shift to γ -Al₂O₃ as adsorbent, the generator capacity can be increased. Simultaneously the activity per mL can also be increased by using smaller bed size for the column and lower volume of eluent.

2.2. ¹⁸⁸Re Based Radiopharmaceuticals

The Radiopharmaceuticals Division of the Bhabha Atomic Research Centre has been carrying out research on the development of ¹⁸⁸Re radiopharmaceuticals for over 25 years. The following sections give details of some of the important developments.

2.3. ¹⁸⁸Re(V)DMSA (DMSA – Dimercapto Succinic Acid)

^{99m}Tc(V)DMSA is a widely used radiopharmaceutical for diagnostic imaging of medullary carcinoma of thyroid [9-11], head and neck tumors, [12, 13], soft tissue tumors and metastatic bone lesions and osseous metastasis from breast cancer [14-16]. Rhenium being the therapeutic analogue of ^{99m}Tc, which shares similar chemistry, envisaging use of ^{186/188}Re(V)DMSA for therapeutic applications is obvious. Bisunadan et al. reported the first study on the preparation and use of ¹⁸⁶Re(V)DMSA for tumor therapy [17]. Subsequently, several others reported the preparation and use of ^{186/188}Re(V)DMSA for different cancers [18-20]. General approach for the preparation of ^{186/188}Re(V)DMSA involves heating DMSA with Na^{186/188}ReO₄ in presence of Sn²⁺ at 100°C. Following this method, Kothari *et al.* reported the preparation of ¹⁸⁶Re(V)DMSA and its biological evaluation in Wistar rats [21]. They observed significant uptake of the radiotracer in kidneys, which limited wide spread application of this therapeutic agent in the clinic. Attempts to reduce the kidney uptake of ^{186/188}Re(V)DMSA using blocking agents were not successful [22]. Later, Kothari et al. found that signifycant reduction in kidney uptake of ^{186/188}Re(V)DMSA could be achieved by modifying the method of preparation of the radiotracer. The ^{186/188}Re(V)DMSA prepared using sodium metabisulphite $(Na_2S_2O_5)$ as reducing agent in place of stannous chloride (SnCl₂), showed significant reduction in kidney uptake (only 0.68±0.06 %ID/g at 24 h post injection (p.i.)). It is pertinent to note that kidney uptake observed with ^{186/188}Re(V)DMSA prepared by conventional method, using stannous chloride as reducing agent, was 2.93±0.93 %ID/g at the same time point [23]. Though the reason for this observation was not understood, authors observed a noticeable change in the isomeric ratio of the ^{186/188}Re(V)DMSA complex. The highperformance liquid chromatography (HPLC) analysis of ^{186/188}Re(V)DMSA prepared by conventional method using stannous chloride as reducing agent showed three major peaks representing anti, syn-endo and syn-

Table 1: Summary of ¹⁸⁸W/¹⁸⁸Re Generators Developed Using Nanosorbents

Sorbent	Dynamic Sorption Capacity (mg/g)	Activity of ¹⁸⁸ W Loaded (GBq)	Elution Yield of ¹⁸⁸ Re (%)	Maximum Radioactive Concentration of ¹⁸⁸ Re (GBq/mL)	Level of ¹⁸⁸ W Impurity in ¹⁸⁸ Re (%)	Radiochemical Purity of ¹⁸⁸ ReO₄ ⁻ (%)	Consistency in Generator Performance
TiP	100	1.85	> 80	0.37	< 10 ⁻³	>99	Consistent performance for 6 months
ZrO ₂	120	1.85	> 78	0.33	< 10 ⁻⁴	>99	
γ -Al ₂ O ₃	326	11.1	> 80	1.85	< 10 ⁻³	>99	

exo isomers, respectively, as shown in Figure 1(A) [24]. Apart from these major peaks, a minor peak representing an unknown structure was observed between the anti and syn-endo isomeric forms. Kothari *et al.* observed significant contribution from this peak in ^{186/188}Re(V)DMSA prepared using sodium metabisulphite reducing agent [Figure 1(B)] [23]. However, it is not clear whether it has any correlation with reduction in kidney uptake and the phenomenon is worth investigating.

Recently, our group developed a lyophilized kit for the preparation of ¹⁸⁸Re(V)DMSA at room temperature [unpublished results]. It is known that presence of oxalate facilitate reduction of rhenium, therefore the room temperature kit (RT kit) contained sodium oxalate in addition to other ingredients of conventional kit, *i.e.* DMSA and stannous chloride. Using this kit ¹⁸⁸Re(V)DMSA could be prepared in >98% yield at room temperature in 15 min. Though HPLC analysis of ¹⁸⁸Re(V)DMSA prepared using RT kit showed similar isomeric peak patterns, the isomeric ratios were different. The ¹⁸⁸Re(V)DMSA prepared following the conventional method, by heating at 100°C for 30 min, had the anti, syn-endo and syn-exo isomers in the ratio 48:39:13, whereas, the ¹⁸⁸Re(V)DMSA prepared using RT kit at room temperature had the isomeric ratios 48:29:23, respectively. It is interesting to note that experimental conditions had effect only on the synendo and syn-exo isomeric ratio and percentage contribution from the anti isomer remained same. This apparently small change in the isomeric ratio of ¹⁸⁸Re(V)DMSA prepared using RT kit, however, had profound influence on the overall biodistribution pattern in Swiss mice bearing fibrosarcoma tumor. With ¹⁸⁸Re(V)DMSA prepared following conventional

method, we observed uptake in bone (15 %ID/g), kidney (3.2 %ID/g) and tumor (2.2 %ID/g) at 60 min p.i. Subsequently, the activity cleared from all organs and at 24 h p.i., the activity in bone, kidney and tumor were 2.8 %ID/g, 1.5 %ID/g and 0.2 %ID/g, respectively. Similarly, biodistribution of ¹⁸⁸Re(V)DMSA prepared using RT kit showed initial uptake of 6 %ID/g, 1.8 %ID/g and 1.4 %ID/g in bone, kidney and tumor, respectively, at 60 min p.i. It could be noted that uptake values in these organs/tissue are relatively lower compared to the distribution of ¹⁸⁸Re(V)DMSA prepared by conventional method at the same time point. However, at 24 h p.i. a significant increase in bone uptake (22 %ID/g) was observed, while the uptake values in kidney (1.2 %ID/g) and tumor (0.4 %ID/g) were similar to that of ¹⁸⁸Re(V)DMSA prepared by conventional method.

In another experiment, ¹⁸⁸Re(V)DMSA prepared using RT kit was heated at 100°C for 10 min. Subsequent HPLC analysis of the sample revealed that isomeric ratios changed from 48:29:23 to 48:38:14, which was very close to that of ¹⁸⁸Re(V)DMSA prepared using conventional kit. This experiment possibly indicated higher thermodynamic stability of syn-endo isomer vis-à-vis syn-exo isomer. Proceeding further, we carried out biodistribution of ¹⁸⁸Re(V)DMSA prepared by RT kit at room temperature and then heating it again at 100°C for 10 min. As expected, the biodistribution results were similar to the one obtained with ¹⁸⁸Re(V)DMSA prepared by conventional method. These results indicate that higher bone uptake observed in the case of ¹⁸⁸Re(V)DMSA prepared using RT kit at room temperature may be due to the syn-exo isomer. This interesting results advocate further investigation to understand the distribution of individual

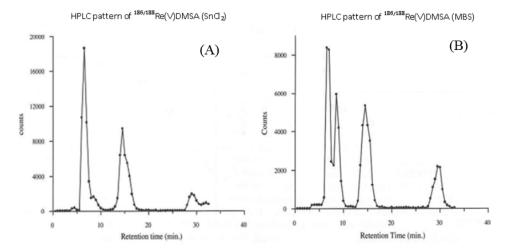


Figure 1: HPLC elution profile of ^{186/188}Re(V)DMSA prepared using different reducing agents (**A**) stannous chloride and (**B**) metabisulphite [23].

International Journal of Nuclear Medicine Research, 2017 71

isomers. If an isomer was found to be suitable for a particular application, modification of reaction conditions to maximize the yield of that isomer could be the next logical step forward.

2.4. ^{186/188}Re-HEDP (HEDP – Hydroxyethane 1,1-Diphosphonic Acid)

 $^{186/188}$ Re-HEDP is a clinically established palliating agent for bone pain due to osseous metastasis [25-31]. Initial HEDP studies were carried out using 186 Re radioisotope. Rhenium-186 is a reactor produced radioisotope from neutron radiative capture (n, γ) reaction on enriched 185 Re or natural rhenium (37.4% enriched in 185 Re).

Kothari et al. reported preparation and evaluated ¹⁸⁶Re-HEDP from ¹⁸⁶Re produced from natural rhenium [32]. This was probably the first study, which used ¹⁸⁶Re produced from natural rhenium in a medium flux (6 x 10¹³ neutrons/cm²/s) reactor. The ¹⁸⁶Re-HEDP prepared under optimized conditions (pH - 2, SnCl₂.2H₂O - 10 mg, HEDP - 50 mg, reaction temperature - 100°C, time - 30 min) was evaluated in normal Wistar rats. The distribution study showed significant uptake of ¹⁸⁶Re-HEDP in bone (30.7 ± 0.04 %ID/organ at 3 h p.i.) and retention thereafter, throughout the duration of the study (29.5 \pm 4.0 %ID/organ at 48 h p.i.). There was no significant uptake of activity in any other major organs. During optimization studies, authors observed that >98% radiochemical purity could be achieved with HEDP amount as low as 1.25 mg/mL, stannous chloride 0.4 mg and total rhenium content 100 µg, rest of the reaction conditions remaining the same. However, to achieve stability of ¹⁸⁶Re-HEDP for extended periods, significantly higher amounts of the reactants were necessary. This observation was in agreement with a study reported by Elder et al., which established the influence of amounts of HEDP, stannous chloride and the total rhenium content on the final structure of Re-HEDP, and consequently, its biodistribution [33]. Use of natural rhenium resulted in, unavoidable, co-production of ¹⁸⁸Re (Half-life – 16.9 h). Therefore, the authors cooled the irradiated sample for four days to reduce the activity of ¹⁸⁸Re to ~6% of the total activity. Authors noticed, and highlighted, the practical difficulty with this approach of ¹⁸⁶Re-HEDP preparation, since it resulted in significant reduction in ¹⁸⁶Re-activity during the cooling period. By this approach, preparation of clinical dose of ¹⁸⁶Re-HEDP was virtually impractical. As an alternative, authors proposed mixed radionuclide therapy with ^{186/188}Re-HEDP, which could eliminate the

4 day cooling requirement. However, dosimetry of mixed radionuclide therapy approach with ^{186/188}Re-HEDP was challenging, which probably limited its clinical applications.

With commercial availability of ¹⁸⁸W/¹⁸⁸Re generator, several studies using ¹⁸⁸Re-HEDP were reported. A comparative clinical study of ¹⁸⁸Re-HEDP, ¹⁸⁶Re-HEDP, ¹⁵³Sm-EDTMP and ⁸⁹Sr by Liepe *et al.* demonstrated similar efficacy of these agents for palliation of pain from osseous metastasis [30]. The authors concluded that efficacy being comparable, choice of therapeutic radionuclide may be decided based on availability of the radioisotope and logistics. In this respect, ¹⁸⁸Re-HEDP enjoyed clear advantage over the other pain-palliating agents based on reactor produced radioisotopes, since it can be prepared *on demand* in any hospital radiopharmacy housing a ¹⁸⁸W/¹⁸⁸Re generator.

The growth of nuclear medicine can be partly attributed to the availability of lyophilized radiopharmaceutical kits and ⁹⁹Mo/^{99m}Tc generator. For a hospital radiopharmacy having a ¹⁸⁸W/¹⁸⁸Re generator, availability of lyophilized kits for the preparation of ¹⁸⁸Re-radiopharmaceuticals is an advantage. Verdera et al. reported the first lyophilized kit for the preparation ¹⁸⁸Re-HEDP [35]. Preparation of ¹⁸⁸Re-HEDP requires addition of carrier rhenium to carrier-free Na¹⁸⁸ReO₄ from ¹⁸⁸W/¹⁸⁸Re generator. This is essential to obtain the ¹⁸⁸Re-HEDP species which shows the pharmacokinetics necessary for a bone pain palliating agent. Along with HEDP (10 mg), gentisic acid (3 mg) and stannous chloride dihydrate (3.7 mg), Verdera et al., included carrier potassium perrhenate (300 μ g, ~1 μ mol) in their lyophilized HEDP kit. Though the authors have not mentioned the shelf-life of such lyophilized HEDP kits, presence of stannous and carrier perrhenate together in the kit could significantly shorten the shelf-life of the kit.

Mallia *et al.* circumvented this problem of short shelf-life of the kit by excluding the carrier perrhenate from the HEDP kits [36]. While rest of the kit components and their amounts remained similar to that of the kit reported by Verdera *et al.*, preparation of ¹⁸⁸Re-HEDP using this kit required addition of 1µmol of carrier perrhenate along with freshly eluted Na¹⁸⁸ReO₄ from ¹⁸⁸W/¹⁸⁸Re generator. Subsequently, heating the kit vial at 100°C for 20 min resulted in the formation of ¹⁸⁸Re-HEDP in more than 98% radiochemical purity. After adjusting the pH of the preparation to physiological conditions (using sterile sodium acetate provided with the kit), the preparation is ready for patient administration. Shelf-life of this kit was one year from the date of production. Clinical evaluation of ¹⁸⁸Re-HEDP prepared using these kits are being carried out in different nuclear medicine centres in India. The development of lyophilized HEDP kit in BARC is well-timed considering the increasing number of nuclear medicine centres in India offering ¹⁸⁸Re-therapy.

2.5. ¹⁸⁸ReN-DEDC/Lipiodol (DEDC – Diethyl Dithiocarbamate)

Hepatocellular carcinoma (HCC) is the most common primary liver cancer and represents the second most common causes of cancer death worldwide [37]. Patients presented with late stage HCC, which are unresectable, are recommended for loco-regional therapies such as transarterial chemoembolization (TACE), selective internal radiation therapy (SIRT) or sorafinib therapy, depending on the stage of the disease [38]. SIRT is also called TARE or transarterial radioembolization. TARE is similar to TACE, the only difference being the use of radiation emitted from radioisotopes for therapy in former, while the latter uses chemotherapeutics. TARE using radiolabelled particles and colloids are one of the options for inoperable HCC patients. There are several options for doing TARE, which include ⁹⁰Y-labeled particulates based on glass (TheraSphere®) or resin (SIR-Spheres[®]) and ¹³¹I- or ¹⁸⁸Re-labeled lipiodol. Especially in India, high cost of ⁹⁰Y-microspheres and non-availability of local substitutes has limited this mode of therapy to a very small group of patients who can afford it.

Wang *et al.* reported the preparation of ¹⁸⁸Relabeled lipiodol for TARE [39]. Their approach involved preparation of ¹⁸⁸Re-EDTB (EDTB - N,N,N",N"tetrakis(2-benzymidazolylmethyl)-I,2-ethanediamine) and subsequent incubation with EDTB-lipiodol at 70°C for 2h. This procedure resulted in ¹⁸⁸Re-lipiodol >98% yield. Biodistribution studies carried out in Sprague Dawley rats showed slow clearance of activity from liver, however, accumulation of significant levels of activity in lungs and kidneys severely limited its clinical applications.

Jeong *et al.* reported preparation and use of a lipiodol solution of ¹⁸⁸Re-TDD (TDD - 2,2,9,9-Tetramethyl-4,7-diaza-1,10-decanedithiol) for therapy of liver cancer [40]. This method involved preparation of a lipophilic complex, ¹⁸⁸Re-TDD, and subsequent extraction into lipiodol. Due to the lipophilic nature of the ¹⁸⁸Re-TDD complex, it is retained in lipiodol phase. This radioactive lipiodol solution was used for TARE procedure. Though ¹⁸⁸Re-TDD could be prepared in good yields and extracted into lipiodol, authors noticed that its retention in liver is not good enough to treat liver cancer patients. The same group subsequently reported a modified form of ¹⁸⁸Re-TDD, ¹⁸⁸Re-HDD (HDD - 4-hexadecyl-2,2,9,9-tetramethyl-4,7-diaza-1,10decanedithiol), which showed higher retention in liver. Kumar *et al.* reported an IAEA-sponsored clinical study, which concluded that TARE with ¹⁸⁸Re-HDD is safe and effective option for the therapy of inoperable hepatocellular carcinoma [41]. Lyophilized kits for the preparation of ¹⁸⁸Re-HDD are available commercially.

Although, ¹⁸⁸Re-HDD complex could be prepared in high yields with commercially available kits, extraction of the complex into the lipiodol phase was not satisfactory. Boschi et al. reported preparation of a lipiodol solution of ¹⁸⁸ReN-DEDC (DEDC – diethyl dithiocarbamate) complex [42]. This approach was very similar to that of Jae Min Jeong et al. wherein a lipophilic ¹⁸⁸Re-complex was prepared and extracted into lipiodol. The complex ¹⁸⁸ReN-DEDC is prepared in two steps. First step involved preparation of ¹⁸⁸Renitrido core, which was subsequently used for radiolabeling DEDC ligand to obtain ¹⁸⁸ReN-DEDC complex. By this method, ¹⁸⁸ReN-DEDC complex could be prepared in >97% radiochemical purity and >96% extraction of the complex into lipiodol could be achieved. Clinical trials with ¹⁸⁸ReN-DEDC/lipiodol showed retention of activity in liver with no activity accumulation seen in lungs, kidneys or any other vital organs.

Considering increasing number of liver cancer incidences in India, a need for locally available, effective and economical TARE agent was felt. Following the procedure reported by Prof. Duatti's group, we developed a two-vial kit for the preparation of ¹⁸⁸ReN-DEDC. First kit vial used for the preparation of ¹⁸⁸ReN core, contained 2 mg of DTCz (N-methyl-Smethyl-dithiocarbazate), 28 mg of sodium oxalate and 0.8 mg of stannous chloride dihydrate. The second kit contained 27 mg of DEDC and carbonate buffer (1 M). The procedure involved addition of up to 3 mL of freshly eluted Na¹⁸⁸ReO₄ containing 150 µL of glacial acetic acid to the kit vial 1. Upon incubating at room temperature for 30 min, ¹⁸⁸ReN core could be prepared in >90% yield. In the second step, kit vial 2 was reconstituted with 2 mL of sterile saline and 1.5 mL of the solution is aseptically transferred to kit vial 1.

Subsequently, kit vial 1 was heated at 70°C for 20 min to obtain ¹⁸⁸ReN-DEDC complex. Analysis of the reaction mixture showed that ¹⁸⁸ReN-DEDC complex was formed in >80% purity. In the third step, 1-2 mL of sterile lipiodol was added to kit vial 1, the contents were thoroughly mixed for 5 min and subsequently centrifuged for 10 min to effect separation of the two layers. The lipiodol layer containing the ¹⁸⁸ReN-DEDC complex was carefully separated and used for TARE. The SPECT/CT images obtained during preliminary clinical trials using ¹⁸⁸ReN-DEDC/lipiodol, carried out in Tata Memorial Hospital, Mumbai, showed retention of the ¹⁸⁸Re-activity in liver 24 h p.i., with no detectable accumulation of activity in lungs or kidneys. A regulatory clearance to manufacture and supply the kits in India is underway.

2.6. ¹⁸⁸Re-Agents for Radiation Synovectomy

Radiation synovectomy is an effective treatment for rheumatoid arthritis [43-46]. This modality, which is an alternative to surgical intervention, involves intraarticular injection of beta emitting radiopharmaceutical to control the synovial inflammation. A critical requirement of radiation synovectomy agents is their ability to localize at the point of injection, since leaching of the agent will render the therapy ineffective. A general approach to minimize the effect of leaching is to use a short-lived radioisotope such that therapeutic dose can be delivered at the point of inflammation in a short span of time. Another approach to minimize leaching is to use radiolabeled particles with larger size [47].

Yttrium-90 and ³²P are two radioisotopes commonly used for the preparation of synovectomy agents [48, 49]. Both radioisotopes have high-energy beta emissions suitable for synovectomy applications. However, being pure beta emitters, both radioisotopes exclude the possibility of imaging to assess localization of the injected radiopharmaceutical. Additionally, the 14 day half-life of ³²P is too long and in the event of leaching it may result in radiation dose to other organs/tissues. In this context, ¹⁸⁸Re has significant advantages over both ⁹⁰Y and ³²P, owing to its favorable physical and chemicals properties described in section 1. Additionally, it has relatively short half-life (16.9 hours) and it is a generator produced radioisotope. Preparation and evaluation of ¹⁸⁸Re labeled microspheres and ¹⁸⁸Re-suphur colloids was reported earlier [50-53]. However, significant leaching from the site of injection severely limited their application. Hydroxyl apatite (HA), a major component

of the bone matrix, had been radiolabeled with ^{186/188}Re for synovectomy applications [54, 55]. However, these agents exhibited poor *in vitro* stability, which limited their application.

Kothari et al. reported direct labeling of HA with ¹⁸⁸Re [47]. Their method involved incubation of freshly eluted Na¹⁸⁸ReO₄ with HA (10 mg), sodium oxalate (10 mg) and stannous chloride (10 mg) at room temperature for 1 hour. The pH of the solution was using hydrochloric maintained at 1.4 acid. Subsequently, the ¹⁸⁸Re-HA particles were washed with ascorbic acid to remove free perrhenate and resuspended in ascorbic acid medium. By this method, ¹⁸⁸Re-HA particles could be prepared in >98% yield. Authors observed that presence of oxalate significantly reduced the dissolution of HA under acidic conditions. Particle size of cold Re-HA, determined by laser diffraction, was found to be between 2-20 μ m (>90%). ¹⁸⁸Re-HA particles were found to be stable in ascorbic acid medium for up to 4 days. Upon intraarticular injection of ¹⁸⁸Re-HA particles in rat joints, extraarticular leakage of only 2.5% was observed even after 2 days post injection. Overall, this study demonstrated the potential of ¹⁸⁸Re-HA particles for therapy of inflamed synovial joints.

2.7. Miscellaneous ¹⁸⁸Re-Radiopharmaceuticals

2.7.1. ¹⁸⁸Re-Skin Patches

Application of radioactive skin patches is an effective mode of therapy for skin cancers and superficial tumors [56]. This modality is an alternative to teletherapy, brachytherapy or kilo voltage x-ray therapy for the treatment of superficial cancers. Papers or skin patches incorporating beta emitting radionuclides such as ¹⁸⁸Re or ¹⁶⁶Ho has been reported earlier [57-59].

Mukharjee *et al.* reported a modified procedure for the preparation of ¹⁸⁸Re-bandage patch [60]. Their method involved preparation of ¹⁸⁸Re-tin colloid followed by trapping of the radioactive colloidal particle on to a 0.22 μ m Millipore filter by passing the solution through the filter. The filter paper was subsequently sandwiched between two nitrocellulose membranes and then placed on an adhesive bandage for therapeutic applications. Radioactive bandages thus prepared were evaluated in melanoma tumor bearing C57BL/6 mice. The study conclusively demonstrated the efficacy and potential of ¹⁸⁸Re-bandages for the therapy of superficial tumors [Figure **2**].

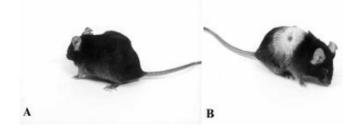


Figure 2: Control mouse (A) bearing melanoma (25 day old tumor); and mouse treated with skin patch application after five months (B) [60].

2.7.2. ¹⁸⁸Re-EC (EC-Ethylene Dicystene) for Endovascular Therapy

Restenosis is an event with significant probability following coronary angioplasty [61, 62]. One of the procedures followed to avoid/minimize restenosis of dilated artery by balloon angioplasty is by local delivery of radiation dose to the dilated arteries [63-65]. Dose delivery is generally achieved using balloon filled with radioactive solution by a procedure similar to balloon angioplasty. One of the inherent risks in this procedure is the rupture of balloon during the procedure leading to leakage of the radioactive liquid into circulation. Therefore, the pharmacokinetics of the radiotracer should be such that in the event of a balloon rupture, the activity should clear from the body within a short period of time, such that dose burden on the patient could be minimized. A comparison between Na¹⁸⁸ReO₄, ¹⁸⁸Re-MAG₃ and ¹⁸⁸Re-DTPA for this application revealed that $^{188}\mbox{Re-MAG}_3$ is most suitable for the purpose [64]. However, the long-term stability of ¹⁸⁸Re-MAG₃ is doubtful [66].

Das et al. [67] reported the preparation of ^{186/188}Re-EC, an analogue of ^{99m}Tc-EC, which is a clinically used renal agent, as a substitute for ¹⁸⁸Re-MAG₃. Optimized procedure for the preparation of ^{186/188}Re-EC involved incubation of a mixture of 10 mM solution of EC with stannous chloride (2 mg) and Na $^{186/188} ReO_4$ at 100°C for 20 min. The pH of the reaction mixture was maintained at 2. By this method, ¹⁸⁸Re-EC could be prepared in >98% radiochemical purity. Biodistribution studies of ^{186/188}Re-EC carried out in Wistar rats indicated that in terms of clearance and dose delivered to other vital organs, ^{186/188}Re-EC had distinct advantages over other rhenium agents used for endovascular brachy therapy applications. In addition, ^{186/188}Re-EC was also found to retain its radiochemical purity for a longer time compared to ¹⁸⁸Re-MAG₃.

2.7.3. ¹⁸⁶Re-CTMP (CTMP – 1,4,8,11-Tetraaza Cyclotetradecyl-1,4,8,11-Tetramethylene Phosphonic Acid) for Bone Pain Palliation

Phosphonates have shown good affinity for bone matrix and they have been utilized to deliver therapeutic dose of radiation to pain causing metastatic or primary bone lesions to obtain palliative effect to the patients. A number of phosphonates radiolabeled with beta emitting radionuclides such as ¹⁸⁶Re, ¹⁸⁸Re, ¹⁵³Sm, ¹⁷⁷Lu etc. have been prepared and evaluated [25-34]. A number of such phosphonate radiopharmaceutials are in clinical use today.

Kothari et al. had earlier reported the preparation and evaluation of ^{186/188}Re-HEDP for bone pain palliation [32]. The same group reported synthesis, radiolabeling and evaluation of a cyclic phosphonate for bone pain palliation [68]. Starting from cyclam (1,4,8,11-tetraaza cyclotetradecane), formaldehyde and orthophosphoric acid, a tetra phosphonate ligand, cyclotetradecyl-1,4,8,11-tetrame-1.4.8.11-tetraaza thylene phosphonic acid or CTMP, was synthesized. This ligand was subsequently radiolabeled with Na¹⁸⁶ReO₄ by heating at 100°C for 30 min in presence of stannous chloride as reducing agent. The pH of the reaction mixture was maintained at 2. By this procedure, ¹⁸⁶Re-CTMP could be prepared in >97% RCP. Authors observed that $^{186}\mbox{Re-CTMP}$ complex remained stable at room temperature for up to 6 days post preparation. Biodistribution of ¹⁸⁶Re-CTMP in normal Wistar rats showed significant uptake and retention of the complex in skeleton with fast clearance from other non-target organs. A comparison with ¹⁸⁶Re-HEDP indicated that the pharmacokinetic parameters of ¹⁸⁶Re-CTMP are either similar or superior to the former [Figure 3]. Overall, ¹⁸⁶Re-CTMP complex demonstrated its potential as a possible bone painpalliating agent.

2.7.4. ^{186/188}Re-Labeled Porphyrin for Targeted Radiotherapy

Porphyrin is a structural unit central to the oxygen transporter in our body, the hematoporphyrin. Porphyrins have shown preferential accumulation in neoplastic tissues, lymph nodes, embryonic and traumatized tissues [69-72]. This property made them potential molecules for diagnosis and therapy of various diseases such as cancer [69-77]. Wong *et al.* reported a ^{99m}Tc-hematoporphyrin for *in vivo* imaging applications [78]. In this complex, ^{99m}Tc was tagged at the periphery of the porphyrin ring. Thereafter, several studies were reported wherein different derivatives of porphyrins were radiolabeled with radioisotopes such

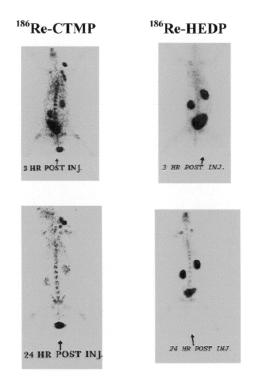


Figure 3: Scintigraphic images of ¹⁸⁸Re-CTMP and ¹⁸⁸Re-HEDP [68].

as ⁵⁷Co, ⁶⁴Cu, ¹⁰⁹Pd etc. [79-81]. A notable difference between the ^{99m}Tc-hematoporphyrin and the ⁵⁷Co/⁶⁴Cu/¹⁰⁹Pd-porphyrins is that in the latter, the metal atom (⁵⁷Co, ⁶⁴Cu or ¹⁰⁹Pd) occupied the centre of the porphyrin ring. However, ⁵⁷Co/⁶⁴Cu/¹⁰⁹Pd-porphyrin derivatives failed to show selective uptake in target tissue. This observation possibly indicates that the porphyrin significantly lose its ability to accumulate in tumor ones a metal atom occupy its core.

Banerjee et al. reported the synthesis of a watermeso-tetrakis[3,4-bis(carboxy soluble porphyrin, methyleneoxy)phenyl] porphyrin, and subsequently radiolabeled it with ^{186/188}Re [82]. Radiolabeling was carried out under basic conditions (pH 9) in presence of stannous tartarate as reducing agent. Upon heating the reaction mixture at 100°C for 30 min, the ^{186/188}Reporphyrin complex could be prepared in >98% yield. Biodistribution studies of ^{186/188}Re-porphyrin complex in Swiss mice bearing fibrosarcoma tumor showed uptake in tumor (~3.5% ID/g), and retention till the duration of the study (24 h). Authors observed no significant accumulation of activity in any other vital organ. Major clearance of activity was through renal route.

CONCLUSIONS

Availability of radionuclides at affordable cost is one of the important considerations for the growth of

International Journal of Nuclear Medicine Research, 2017 75

nuclear medicine. The initial investments needed to start a program and adequate revenue returns from it are prime considerations. Considering relatively high cost of ¹⁸⁸W/¹⁸⁸Re generator it is important to use the generator for as many applications as possible. ¹⁸⁸Re-radiopharmaceutical improving Therefore. portfolio is highly essential. Despite the fact that the rhenium radiopharmaceuticals development program was initiated at the Bhabha Atomic Research Centre in the early nineties, clinical utility was limited due to the lack of interest shown by nuclear medicine departments. A new fillip to the program was provided by the entry of Kovai Institute of Medical Sciences (KMCH) which started the routine clinical use of ¹⁸⁸W/¹⁸⁸Re generator for the treatment of HCC and a collaborative program between BARC and KMCH. One of the newer developments thanks to this collaboration is the development of lyophilized DEDC kits for the preparation of ¹⁸⁸ReN-DEDC/lipiodol for the therapy of inoperable HCC. Along with lyophilized DEDC kits, other lyophilized kits for the preparation of ¹⁸⁸Re-HEDP, ¹⁸⁸Re-DMSA etc. will help in effective utilization of ¹⁸⁸Re-activity from ¹⁸⁸W/¹⁸⁸Re generator. The authors also feel it appropriate that the commercial manufacturers explore the utility of high capacity adsorbents discussed in this paper for making generators with higher load of 188W which will provide high activity per mL for the eluted perrhenate solution.

REFERENCES

- Singh, B, Viggars, DA. Nuclear data sheets for A = 188. Nuclear Data Sheets 1981; 33(2): 275-387. https://doi.org/10.1016/S0090-3752(81)80025-7
- [2] Dash A, Chakravarty R. Pivotal role of separation chemistry in the development of radionuclide generators to meet clinical demands, RSC Adv 2014; 4: 42779-42803. <u>https://doi.org/10.1039/C4RA07218A</u>
- [3] Jeong JM, Knapp FF Jr. Use of the Oak Ridge National Laboratory tungsten-188/rhenium-188 generator for preparation of the rhenium-188 HDD/lipiodol complex for trans-arterial liver cancer therapy. Semin Nucl Med 2008; 38(2): S19-29. https://doi.org/10.1053/i.semnuclmed.2007.10.003
- [4] Chakravarty R, Dash A, Venkatesh M. Separation of Clinical Grade 188Re from 188W Using Polymer Embedded Nanocrystalline Titania 2009; 69(11): 1363-1372.
- [5] Chakravarty R, Shukla R, Tyagi AK, Dash A, Venkatesh M. Nanocrystalline zirconia: a novel sorbent for the preparation of (188)W/(188)Re generator. Appl Radiat Isot 2010; 68(2): 229-238. https://doi.org/10.1016/j.apradiso.2009.10.031
- [6] Chakravarty R, Shukla R, Ram R, Venkatesh M, Tyagi AK, Dash A. Exploitation of nano alumina for the chromatographic separation of clinical grade 188Re from 188W: a renaissance of the 188W/188Re generator technology. Anal Chem 2011; 83(16): 6342-6348. <u>https://doi.org/10.1021/ac201232m</u>
- [7] Chakravarty R, Dash A. Nano Structured Metal Oxides as Potential Sorbents for 188W/188Re Generator: A

Comparative Study. Sep Sci Technol 2013; 48: 607-616. https://doi.org/10.1080/01496395.2012.713433

- [8] British Pharmacopoeia Commission (2008) British pharmacopoeia. The Stationery Office, Norwich. www.pharmacopoeia.org.uk
- [9] Clarke SEM, Lazarus CR, Wraight P, Sampson C, Maisey MN. Pentavalent [^{99m}Tc]DMSA, [¹³¹]]MIBG, and [^{99m}Tc]MDP-An Evaluation of Three Imaging Techniques in Patients with Medullary Carcinoma of the Thyroid. J Nucl Med 1988; 29: 33-38.
- [10] Ohta H, Yamamoto K, Endo K, Mori T, Hamanaka D, Shimazu A, Ikekubo K, Makimoto K, Iida Y, Konishi J, *et al.* A new imaging agent for medullary carcinoma of the thyroid. J Nucl Med 1984; 25(3): 323-325.
- [11] Yen TC, King KL, Yang AH, Liu RS, Yeh SH. Comparative radionuclide imaging of metastatic insular carcinoma of the thyroid: value of technetium-99m-(V)DMSA. J Nucl Med 1996; 37(1): 78-80.
- [12] Ohta H, Endo K, Fujita T, Konishi J, Torizuka K, Horiuchi K, Yokoyama A. Clinical evaluation of tumour imaging using 99Tc(V)m dimercaptosuccinic acid, a new tumour-seeking agent. Nucl Med Commun 1988; 9(2): 105-116.
- [13] Watkinson JC, Lazarus CR, Mistry R, Shaheen OH, Maisey MN, Clarke SE. Technetium-99m (v) dimercaptosuccinic acid uptake in patients with head and neck squamous carcinoma: experience in imaging. J Nucl Med 1989; 30(2): 174-180.
- [14] Babbar A, Kashyap R, Chauhan UP. A convenient method for the preparation of 99mTc-labelled pentavalent DMSA and its evaluation as a tumour imaging agent. J Nucl Biol Med 1991; 35(2): 100-104.
- [15] Chauhan UP, Babbar A, Kashyap R, Prakash R. Evaluation of a DMSA kit for instant preparation of 99mTc(V)-DMSA for tumour and metastasis scintigraphy. Int J Rad Appl Instrum B 1992; 19(8): 825-830. https://doi.org/10.1016/0883-2897(92)90168-X
- [16] Kashyap R, Babbar A, Sahai I, Prakash R, Soni NL, Chauhan UP. Tc-99m(V) DMSA imaging. A new approach to studying metastases from breast carcinoma. Clin Nucl Med 1992; 17(2): 119-122. <u>https://doi.org/10.1097/00003072-199202000-00011</u>
- [17] Bisunadan MM, Blower PJ, Clarke SE, Singh J, Went MJ. Synthesis and characterization of [¹⁸⁶Re]rhenium(V)dimercaptosuccinic acid: a possible tumour radiotherapy agent. Int J Rad Appl Instrum A 1991; 42(2): 167-1671. https://doi.org/10.1016/0883-2889(91)90068-C
- [18] Blower PJ, Kettle AG, O'Doherty MJ, Coakley AJ, Knapp FF Jr. (99m)Tc(V)DMSA quantitatively predicts (188)Re(V)DMSA distribution in patients with prostate cancer metastatic to bone. Eur J Nucl Med 2000; 27(9): 1405-1409. <u>https://doi.org/10.1007/s002590000307</u>
- [19] García-Salinas L, Ferro-Flores G, Arteaga-Murphy C, Pedraza-López M, Hernández-Gutiérrez S, Azorin-Nieto J. Uptake of the 188Re(V)-DMSA complex by cervical carcinoma cells in nude mice: pharmacokinetics and dosimetry. Appl Radiat Isot 2001; 54(3): 413-418. <u>https://doi.org/10.1016/S0969-8043(00)00278-5</u>
- [20] Park JY, Lee TS, Choi TH, Cheon GJ, Choi CW, Awh OD. A comparative study of 188Re(V)-meso-DMSA and 188Re(V)rac-DMSA: preparation and *in vivo* evaluation in nude mice xenografted with a neuroendocrine tumor. Nucl Med Biol 2007; 34(8): 1029-1036. <u>https://doi.org/10.1016/j.nucmedbio.2007.06.016</u>
- [21] Kothari K, Pillai MRA, Unni PR, Shimpi HH, Noronha OP, Samuel AM. Preparation of [¹⁸⁶Re]Re-DMSA and its biodistribution studies. Appl Radiat Isot 1999; 51(1): 43-49. <u>https://doi.org/10.1016/S0969-8043(98)00194-8</u>
- [22] Houston S, Allen S, Lazarus CR, Reghebi K, Blower P, Singh J, Rubens RD, Clarke SEM. Modifying renal uptake of

pentavalent 99Tcm-DMSA and pentavalent 186Re-DMSA offers potential for tumour-targeted radiotherapy in medullary thyroid carcinoma (MTC). Nucl Med Commun 1992; 13: 211. https://doi.org/10.1097/00006231-199204000-00016

- [23] Kothari K, Satpati D, Mukherjee A, Sarma HD, Venkatesh M, Pillai MRA. Kidney uptake of ^{186/188}Re(V)-DMSA is significantly reduced when the reducing agent is changed from stannous ion to metabisulfite. J Label Compd Radiopharm 2002; 45: 675-686. <u>https://doi.org/10.1002/jlcr.598</u>
- [24] Singh J, Reghebi K, Lazarus CR, Clarke SEM, Callahan AP, Knapp FF Jr., Blower PJ. Studies on the preparation and isomeric composition of 186Re- and 188Re-pentavalent rhenium dimercaptosuccinic acid complex. Nucl Med Commun 1993; 14: 197-203. https://doi.org/10.1097/00006231-199303000-00009
- [25] Mathieu L, Chevalier P, Galy G, Berger M. Preparation of Rhenium-186 labelled EHDP and its possible use in the treatment of osseous neoplasms. Int J Appl Radiat Isot 1979; 30: 725-727. https://doi.org/10.1016/0020-708X(79)90150-9
- [26] Maxon HR. 3rd, Schroder LE, Washburn LC, Thomas SR, Samaratunga RC, Biniakiewicz D, et al. Rhenium-188(Sn)HEDP for treatment of osseous metastases. J Nucl Med 1998; 39(4): 659-663.
- [27] Lam MG, de Klerk JM, van Rijk PP. ¹⁸⁶Re-HEDP for metastatic bone pain in breast cancer patients. Eur J Nucl Med Mol Imaging 2004; 31(Suppl 1): S162-S170. https://doi.org/10.1007/s00259-004-1539-4
- [28] Maxon HR, Deutsch EA, Thomas SR, Libson K, Lukes SJ, Williams CC, Ali S. ¹⁸⁶Re-(Sn) HEDP for treatment of multiple metastatic foci in bone: human biodistribution and dosimetric studies. Radiology 1988; 166(2): 501-507. https://doi.org/10.1148/radiology.166.2.3122267
- [29] Maxon HR 3rd, Schroder LE, Hertzberg VS, Thomas SR, Englaro EE, Samaratunga R, Smith H, Moulton JS, Williams CC, Ehrhardt GJ *et al.* Rhenium-186(Sn)HEDP for treatment of painful osseous metastases: results of a double-blind crossover comparison with placebo. J Nucl Med 1991; 32(10): 1877-1881.
- [30] Liepe K, Kotzerke J. A comparative study of ¹⁸⁸Re-HEDP, ¹⁸⁶Re-HEDP, ¹⁵³Sm-EDTMP and ⁸⁹Sr in the treatment of painful skeletal metastases. Nucl Med Commun 2007; 28(8): 623-630. <u>https://doi.org/10.1097/MNM.0b013e32825a6adc</u>
- [31] Palmedo H, Guhlke S, Bender H, Sartor J, Schoeneich G, Risse J, Grünwald F, Knapp FF Jr, Biersack HJ. Dose escalation study with rhenium-188 hydroxyethylidene diphosphonate in prostate cancer patients with osseous metastases. Eur J Nucl Med 2000; 27(2): 123-130. https://doi.org/10.1007/s002590050017
- [32] Kothari K, Pillai MRA, Unni PR, Shimpi HH, Noronha OP, Samuel AM. Preparation, stability studies and pharmacological behavior of [186Re]Re-HEDP. Appl Radiat Isot 1999; 51(1): 51-58. https://doi.org/10.1016/S0969-8043(98)00195-X
- [33] Elder RC, Yuan J, Helmer B, Pipes D, Deutsch K, Deutsch E. Studies of the Structure and Composition of Rhenium-1,1-Hydroxyethylidenediphosphonate (HEDP) Analogues of the Radiotherapeutic Agent (186)ReHEDP. Inorg Chem 1997; 36(14): 3055-3063 https://doi.org/10.1021/ic960980h
- [34] Thapa P, Nikam D, Das T, Sonawane G, Agarwal JP, Basu S. Clinical Efficacy and Safety Comparison of 177Lu-EDTMP with 153Sm-EDTMP on an Equidose Basis in Patients with Painful Skeletal Metastases. J Nucl Med 2015; 56(10): 1513-1519. http://doi.org/10.0067/jjuurged.115.1565762

https://doi.org/10.2967/jnumed.115.155762

[35] Verdera ES, Gaudiano J, León A, Martinez G, Robles A, Savio E, León E, McPherson D W, Knapp FF(Russ) Jr. Rhenium-188-HEDP kit formulation and quality control. Radiochimica Acta 1997; 79: 113. https://doi.org/10.1524/ract.1997.79.2.113

- [36] Mallia MB, Shinto AS, Kameswaran M, Kamaleshwaran KK, Kalarikal R, Aswathy KK, Banerjee S. A Freeze-Dried Kit for the Preparation of (188)Re-HEDP for Bone Pain Palliation: Preparation and Preliminary Clinical Evaluation. Cancer Biother Radiopharm 2016; 31(4): 139-144. https://doi.org/10.1089/cbr.2016.2030
- [37] Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-E386. <u>https://doi.org/10.1002/ijc.29210</u>
- [38] European Association For The Study Of The Liver; European Organisation For Research And Treatment Of Cancer. EASL-EORTC clinical practice guidelines: management of hepatocellular carcinoma. J Hepatol 2012; 56: 908-943, Maida M, Orlando E, Cammà C, Cabibbo G. Staging systems of hepatocellular carcinoma: a review of literature. World J Gastroenterol 2014; 20: 4141-4150.
- [39] Shyh-Jen Wang, Wang-Yu Lin, Min-Nan Chen, Bor-Tsung Hsieh, Lie-Hang Shen, Zei-Tsan Tsai, Gann Ting, F.F.(Russ) Knapp, Radiolabelling of lipiodol with generator-produced 188Re for hepatic tumor therapy, Appl Radiat Isot 1996; 47(3): 267-271.
- [40] Jae Min Jeong, Young Joo Kim, Yoon Sang Lee, Jun II Ko, Miwon Son, Dong Soo Lee, June-Key Chung, Jae Hyung Park, Myung Chul Lee, Lipiodol solution of a lipophilic agent, 188Re-TDD, for the treatment of liver cancer, Nucl Med Biol 2001; 28(2): 197-204. <u>https://doi.org/10.1016/S0969-8051(00)00208-0</u>
- [41] Kumar A, Srivastava DN, Chau TT, Long HD, Bal C, Chandra P et al. Inoperable hepatocellular carcinoma: transarterial 188Re HDD-labeled iodized oil for treatment--prospective multicenter clinical trial. Radiology 2007; 243(2): 509-519. https://doi.org/10.1148/radiol.2432051246
- [42] Boschi A, Uccelli L, Duatti A, Colamussi P, Cittanti C, Filice A, et al. A kit formulation for the preparation of 188Relipiodol: preclinical studies and preliminary therapeutic evaluation in patients with unresectable hepatocellular carcinoma. Nucl Med Commun 2004; 25(7): 691-699. https://doi.org/10.1097/01.mnm.0000130241.22068.45
- [43] Deutsch E, Brodack KF, Deutsch KF. Radiation synovectomy revisited. Eur. J Nucl Med 1993; 20: 1113-1117. https://doi.org/10.1007/BF00173494
- [44] Doherty M. Potential rheumatologic applications for intraarticular radiocolloid therapy. Geriatric Med Today 1984; 3: 31-42.
- [45] Hosain F, Haddon MJ, Hosain H, Drost JK, Spencer RP. Radiopharmaceuticals for diagnosis and treatment of arthritis. Nucl Med Biol 1990; 17: 151-155. <u>https://doi.org/10.1016/0883-2897(90)90017-u</u>
- [46] Modder G. 1995. Radiosynoviorethesis-Involvement of Nuclear Medicine in Rheumatology and Orthopedics. Warlich Druck and Verlagsges, Meckenheim, Germany, Rosenthall L, 1978. Use ofradiocolloids for intra-articular therapy for synovitis. In: Spencer, R.P. (Ed.), Therapy in Nuclear Medicine. Grune and Stratton, New York, pp. 147-153.
- [47] Kothari K, Suresh S, Sarma HD, Meera V, Pillai MRA. 188Re-labeled hydroxyapatite particles for radiation synovectomy. Appl Radiat Isot 2003; 58(4): 463-468. https://doi.org/10.1016/S0969-8043(03)00028-9
- [48] Bowen BM, Darracott J, Garnett ES, Tomlinson RH. Yttrium 90 citrate colloid for radioisotope synovectomy. Am. J Hosp Pharm 1975; 32: 1027-1030.
- [49] Pandey U, Mukherjee A, Chaudhary PR, Pillai MRA, Venkatesh M. Preparation and studies with 90Ylabelled particles for use in radiation synovectomy. Appl Radiat Isot

2001; 55: 471-475. https://doi.org/10.1016/S0969-8043(01)00061-6

- [50] Venkatesan P, Shortkroff S, Zalutsky MR, Sledge CB. Rhenium heptasulfide: a potential carrier system for radiation synovectomy. Nucl Med Biol 1990; 4: 357-362. <u>https://doi.org/10.1016/0883-2897(90)90101-6</u>
- [51] Wang SJ, Lin WY, Hsieh BT, Shen LH, Tsai ZT, Ting G, Knapp Jr. FF, 1995. Rhenium sulfur colloid as a radiation synovectomy agent. Eur. J Nucl Med 1995; 22: 505-507. <u>https://doi.org/10.1007/BF00817272</u>
- [52] Wang SJ, Lin WY, Chen MN, Hsieh BT, Shen LH, Tsai ZT, et al. Rhenium 188 microspheres: a new radiation synovectomy agent. Nucl Med Comm 1998; 19: 427-433. <u>https://doi.org/10.1097/00006231-199805000-00004</u>
- [53] Wang SJ, Lin WY, Chen MN, Chen JT, Ho WL, Hsieh BT, et al. Histologic study of effects of radiation synovectomy with rhenium 188 microspheres. Nucl Med Biol 2001; 28: 727-732.

https://doi.org/10.1016/S0969-8051(01)00228-1

- [54] Chinol M, Vallabhajosula S, Goldsmith SJ, Klein MJ, Deutsch KF, Chinen LK, *et al.* Chemistry and biological behavior ofsamarium-1 53 and rhenium-186-labeled hydroxyapatite particles: potential radiopharmaceuticals for radiation synovectomy. J Nucl Med 1993; 34: 1536-1542.
- [55] Grillenberger KG, Glatz S, Reske SN. Rhenium –188 labeled hydroxyapatite and rhenium–188 sulfur colloid. *In vitro* comparison oftwo agents for radiation synovectomy. Nuklearmedizin 1997; 36: 71-75.
- [56] Mukherjee A, Pandey U, Sarma HD, Pillai MRA, Venkatesh M. Preparation and evaluation of ⁹⁰Y-skin patches for therapy of superficial tumors in mice. Nuclear Medicine Communication 2002; 23: 243-247. <u>https://doi.org/10.1097/00006231-200203000-00007</u>
- [57] Lee JD, Park KK, Lee MG, Kim EH, Rhim KJ, Lee JT, et al. Radionuclide therapy of skin cancers and Bowen's disease using a specially designed skin patch. Journal of Nuclear Medicine 1997; 38(5): 697-702.
- [58] Chung YL, Lee JD, Bang D, Lee JB, Park KB, Lee MG. Treatment of Bowen's disease with a specially designed radioactive skin patch. European Journal of Nuclear Medicine 2000; 27: 842-846. https://doi.org/10.1007/s002590000262
- [59] Jeong JM, Lee YJ, Kim EH, Lim SM, Lee DS, Chung JK, Lee MC, Koh CS. Simple preparation of beta ray emitting paper for treatment of skin cancer. Journal of Nuclear Medicine 1998; 39: 234P (abstr).
- [60] Mukherjee A, Pandey U, Sarma HD, Gupta SK, Ingle AD, Pillai MRA, Venkatesh M. Bioevaluation of radioactive bandages in a murine model of melanoma. Int J Radiat Biol 2003; 79(10): 839-845. https://doi.org/10.1080/09553000310001610989
- [61] Hafeli, UO, Lee, EJ, Ciezki, J, Gayle, JP, Martin, BS, Weinhous, MS. Suitability of beta-emitting rhenium for inhibiting restenosis in coronary arteries. J Brachytherapy Intl 1999; 15: 1-11.
- [62] Waksman R. Clinical trials in radiation therapy. Vascular Brachytherapy Monitor 1998; 1: 10-18.
- [63] Knapp FF Jr., Guhlke S, Beets AL, Amols H, Weinberger J. Intraarterial irradiation with rhenium-188 for inhibition of restenosis by PTCA – Strategy and evaluation of species for rapid urinary excretion (abstract). J Nucl Med 1997; 38: 124P.
- [64] Oh SJ, Moon DH, Park SW, Hong MK, Park SJ, Shin JW, et al. Comparison of radiation absorbed dose of Re-188 radiopharmaceuticals for intracoronary radiation therapy in case of balloon rupture (abstract). J Nucl Med 1999; 40: 41P.
- [65] Weinberger J and Knapp FF. Use of liquid-filled balloons for coronary irradiation. In: Vascular Brachytherapy, 2nd ed.

- [66] Wang TS, Weinberger J, Fawwaz RA, Van Heertum RL. Invitro stability evaluation of Re-188-MAG₃: A potential therapeutic agent for prevention of restenosis after Percutaneous Transluminal Coronary Angioplasty. Ann Nucl Med Sci 2001; 14: 19-25.
- [67] Das T, Banerjee S, Samuel G, Kothari K, Unni PR, Sarma HD, et al. [(186/188)Re] rhenium-ethylene dicysteine (Re-Ec): preparation and evaluation for possible use in endovascular brachytherapy. Nucl Med Biol 2000; 27(2): 189-197. https://doi.org/10.1016/S0969-8051(99)00097-9
- [68] Kothari K, Samuel G, Banerjee S, Unni PR, Sarma HD, Chaudhari PR, et al. ¹⁸⁶Re-1,4,8,11-tetraaza cyclotetradecyl-1,4,8,11-tetramethylene phosphonic acid: A novel agent for possible use in metastatic bone-pain palliation. Nucl Med Biol 2001; 28(6): 709-717. https://doi.org/10.1016/S0969-8051(01)00224-4
- [69] Policard A. Etudes sur les aspects offerts par des tumeurs experimentales examinees a la lumiere des woods. C R Soc Biol 1924; 91: 14-32.
- [70] Figge FHJ, Weiland GS, Manganiello LOJ. Cancer detection and therapy: Affinity of neoplastic, embryonic and traumatized tissues for porphyrin and metalloporphyrin. Proc Soc Exp Biol Med 1948; 68: 640-641. https://doi.org/10.3181/00379727-68-16580
- [71] Manganiello LOJ, Figge FHJ. Cancer detection and therapy II: methods of preparation and biological effects of metalloporphyrin. Bull School Med Univ Maryland 1951; 36: 3-7.
- [72] Peck GC, Mack HP, Figge FHJ. Cancer detection and therapy III: Affinity of lymphatic tissues for hematoporphyrin. Bull School Med Univ Maryland 1953; 38: 124-127.
- [73] Rasmussen-Taxdal DS, Ward GE, Figge FHJ. Fluorescence of human lymphatic and cancer tissues following high doses of intravenous hematoporphyrin. Cancer 1955; 8: 78-81. <u>https://doi.org/10.1002/1097-0142(1955)8:1<78::AID-CNCR2820080109>3.0.CO:2-L</u>

Received on 26-04-2017

Accepted on 04-05-2017

Published on 31-07-2017

http://dx.doi.org/10.15379/2408-9788.2017.06

© 2017 Mallia et al.; Licensee Cosmos Scholars Publishing House.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/), which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.

- [74] Altman KF, Solomon K. Localization of a halogenated porphyrin in mouse tumour. Nature 1960; 187: 1124. <u>https://doi.org/10.1038/1871124a0</u>
- [75] Winkelman J. Intracellular localization of hematoporphyrin in a transplanted tumour. J Natl Cancer Inst 1961; 27: 1369-1377.
- [76] Lipson RL, Baldes EJ, Gray MS. Hematoporphyrin derivative for detection and management of cancer. Cancer 1967; 20: 2255-2257. <u>https://doi.org/10.1002/1097-0142(196712)20:12<2255::AID-CNCR2820201229>3.0.CO:2-U</u>
- [77] Fawwaz RA, Wang TST, Alderson PO. Evaluation of radioporphyrins as tumour seeking agents in experimental animals. J Nucl Med 1981; 22: P50 (abstract).
- [78] Wong DW. A simple chemical method of labeling hematoporphyrin derivative with Technetium-99m. J Label Comp Radiopharm 1982; 20: 351-361. https://doi.org/10.1002/ilcr.2580200305
- [79] Hambright P, Fawwaz R, Valk P, McRae J, Bearden AJ. The distribution of various water soluble radioactive metalloporphyrins in tumor bearing mice. Bioinorg Chem 1975; 5(1): 87-92. <u>https://doi.org/10.1016/S0006-3061(00)80224-0</u>
- [80] Hambright P, Smart JC, McRae J, Nohr ML, Yano Y, Chu P, Bearden AJ. Tumor imaging with 57-cobalt (III) sandwich complexes and 57-cobalt (III) porphyrins. Inorg Nucl Chem Lett 1976; 17: 217-222. https://doi.org/10.1016/0020-1650(76)80200-0
- [81] Zanelli GD, Kaelin AD. Synthetic porphyrins as tumour localizing agents. Br J Radiol 1981; 54: 403-407. <u>https://doi.org/10.1259/0007-1285-54-641-403</u>
- [82] Banerjee S, Das T, Samuel G, Sarma HD, Venkatesh M, Pillai MRA. A novel [186/188Re]-labelled porphyrin for targeted radiotherapy. Nucl Med Commun 2001; 22(10): 1101-1107. https://doi.org/10.1097/00006231-200110000-00008